

ГОДИШНИК НА СОФИЙСКИЯ УНИВЕРСИТЕТ „СВ. КЛИМЕНТ ОХРИДСКИ“

БИОЛОГИЧЕСКИ ФАКУЛТЕТ

Книга 2 – Ботаника

Том 102, 2018

ANNUAL OF SOFIA UNIVERSITY “ST. KLIMENT OHRIDSKI”

FACULTY OF BIOLOGY

Book 2 – Botany

Volume 102, 2018

ANTIOXIDANT POTENTIAL OF BULGARIAN YARROW AND THYME

NELLY G. GEORGIEVA, GANKA T. CHANEVA & MIROSLAVA K. ZHIPONOVA*

Department of Plant Physiology, Faculty of Biology, Sofia University “St. Kliment Ohridski”, 8 Dragan Tsankov Blvd., 1164 Sofia, Bulgaria

Abstract. Phenols (sometimes called phenolics) are synthesized by plants for their general defense and particularly in response to antioxidative stress. These biologically active compounds are well known to have beneficial effects on human health. The aim of the present study was to apply a fast in vitro approach to compare the antioxidant potential of yarrow (*Achillea millefolium* L.) and thyme (*Thymus vulgaris* L.) that were freshly collected from their natural habitat in the Rodopi Mts with some randomly selected herbs for infusion from traders in Sofia region. Ethanol extracts from yarrow and thyme plants were compared with an extract from the Chinese white tea (*Camellia sinensis* (L.) Kuntze) used for infusion and known for its high phenolic content with antioxidant effect. The total quantity of phenols in both studied herbs varied in a close range but was 5-8 times lower than this in the white tea. The average antioxidant activity in thyme was slightly higher than in yarrow but was nearly four times lower in comparison to their activity in the white tea. Slight variations between the herbs from the Rodopi Mts and Sofia were found in the phenolic content of yarrow and thyme, and in the antioxidant activity of thyme. Significant positive correlation between the content of phenolic and antioxidant activity was observed in white tea and thyme, but not in yarrow but our in vitro approach would need to be confirmed by further in vivo analyses. Our results show that the genotype, habitat and storage conditions could influence the plant antioxidant potential and that it is likely that the Bulgarian herbs contain additional classes of metabolites which determine distinct biological activities.

Key words: antioxidant activity, herbs, phenolics, tea, white tea

*corresponding author: M. K. Zhiponova, Sofia University “St. Kliment Ohridski”, Faculty of Biology, Department of Plant Physiology, 8 Dragan Tsankov Blvd., BG-1164, Sofia, Bulgaria; zhiponova@uni-sofia.bg

INTRODUCTION

The secondary plant metabolites serve as reducing agents that result in decrease of the amount of active oxygen species inside the cell, which prevents further damages and cell malfunction (GUPTA & SHARMA 2006; PEREIRA ET AL. 2013). The phenols (sometimes called phenolics) are secondary metabolites, widespread in herbal plants, and often studied for their antioxidant potential. The variable biological properties of the phenolics are due to their molecular structure including at least one phenol ring in which the hydrogen is usually replaced by a more active residue, such as hydroxyl, methyl or acetyl. These compounds often contain more phenolic rings, therefore they are called polyphenols (DZIALO ET AL. 2016). Plants synthesize phenolic compounds mainly to protect themselves against unfavourable environmental conditions such as ultraviolet light, herbivores and pathogens, as well as to attract pollinators and animals dispersing the seeds (BOUDET 2007). The defence costs are paid mainly in the form of energy, carbon, and nitrogen, while phenolics are suggested to be cheaper than alkaloids because of the additional effort that is required to make inorganic nitrogen bioavailable (MITHÖFER & BOLAND 2012).

Polyphenols are the subject of increasing scientific interest as they have various functions in the human body - antioxidant protection, anti-viral, antibacterial, anti-tumor and anti-inflammatory activity (PANDEY ET AL. 2009). Epidemiological studies and associated meta-analyses strongly suggested that long term consumption of diets rich in plant polyphenols offered some protection against development of cancers, cardiovascular diseases, diabetes, osteoporosis, and neurodegenerative diseases (PANDEY ET AL. 2009). The teas made from *Camellia sinensis* (L.) Kuntze contain polyphenols and flavonoids (mainly catechins), which are considered as their most important phytochemicals in terms of health benefits due to their ability to act as antioxidants by donating electrons or hydrogen protons to reactive oxygen or nitrogen species (SHANNON ET AL. 2018). White teas have been reported to possess higher antielastase, anticollagenase, and antioxidative activity than green tea, which has led to an increased interest in this tea type (THRING ET AL. 2009). In comparison, tisanes derived from herbs or fruit infusions, as chamomile and berry/hibiscus, also contain polyphenols but at significantly lower levels than *C. sinensis* derived teas.

Ethnobotanical studies highlighted the members of the Asteraceae and Lamiaceae families among the most popular medicinal plants in Bulgaria and other countries on the Balkan Peninsula, with different species of yarrow (*Achillea* L.) and thyme (*Thymus* L.) as commonly used herbs (e.g. EVSTATIEVA ET AL. 2007; JARIC ET AL. 2015). The application of these medicinal plants in food industry, cosmetology and pharmacology has been increasingly studied (MEKINIC ET AL. 2014; BOUTAOUI ET AL. 2018). Previous research work on these plants has been mainly confined to their essential oil, however, much attention has recently been

directed to the water-soluble components (BENETIS ET AL. 2008; KRATCHANOVA ET AL. 2010; MEKINIĆ ET AL. 2014; ROGOVA ET AL. 2015; BOUTAOUI ET AL. 2018). In yarrow, phenolic compounds such as flavonoids (*e.g.* vicenin-2, luteolin-3',7-di-O-glucoside, luteolin-7-O-glucoside, rutin, apigenin-7-O-glucoside, luteolin, apigenin) and phenolic acids (*e.g.* rosmarinic acid, m-hydroxybenzoic acid, o-coumaric acid, caffeic, ferulic acid) constitute one of the most important groups of pharmacologically active substances (BENETIS ET AL. 2008; MEKINIĆ ET AL. 2014). Recent investigation on bioactive substances in thyme species also revealed high content of phenolic compounds (benzoic acid, epicatechin, chlorogenic acid, syringic acid, naringin, catechin, o-coumaric acid) - BOUTAOUI ET AL. (2018).

Plant extracts made with water are nutritionally more relevant since herbs are traditionally ingested as hot-water infusions. However, stronger polar solvents (methanol, acetone, ethanol) are preferred for more exhaustive extraction of polyphenol compounds due to their polar groups (BENETIS ET AL. 2008; KRATCHANOVA ET AL. 2010). KRATCHANOVA ET AL. (2010) investigated the influence of the extraction agent on the extractability of polyphenol components and the antioxidant activity of 25 Bulgarian medicinal plants, among which *A. millefolium* and *T. vulgaris*. It was found that the antioxidant potential was higher for 80% acetone extraction than for water extraction.

The phytochemical composition of medicinal plants is influenced by variables such as cultivar, ontogenetic factors, growth conditions, processing conditions, storage (FIEHN 2002; KAPCHINA ET AL. 2014; BOUTAOUI ET AL. 2018; SHANNON ET AL. 2018). In the present study, we applied a simple preliminary, but fast *in vitro* approach to examine the total quantity of phenolic compounds and antioxidant activities of yarrow and thyme herbs from two different regions in Bulgaria, and compared them to the antioxidant potential of Chinese white tea, when using ethanol as a solvent.

MATERIALS AND METHODS

Plant material

The plant material was collected on 24th June 2017, from a natural habitat in Bulgaria, the Rodopi Mts (Plovdiv Province, village Dryanovo, latitude 41.7946091; longitude: 24.7867012; altitude 1000 m a.s.l.). The voucher specimens were deposited in the Herbarium of Sofia University „St. Kliment Ohridski“, as follows: SO107842 for *Achillea millefolium* L. (yarrow) and SO107844 for *Thymus vulgaris* L. (thyme). Flowers were air dried at room temperature in darkness until no significant change of the dry weight was detected. The samples were analyzed four weeks after the collection. The commercially purchased yarrow and thyme herbs were randomly selected from Bulgarian producers in Sofia region (at average altitude of 500 m a.s.l.). The white tea (*Camellia sinensis*) consisted of unopened buds and it was purchased from a herbal pharmacy in Sofia in 2017.

Preparation of extracts

For extract preparation 50 mg of air-dried plant material was homogenized with 5 ml 100% ethanol and disintegrated in ultrasonic bath for 2 min. After centrifugation (at 9000 rpm for 20 min) the supernatant was subjected to further analyses.

Total phenolic content analysis

The total phenolic content was determined according to SINGLETON ET AL. (1999). Test samples contained 0.1 ml plant extract, 1.5 ml Folin-Ciocalteu reagent (previously dissolved in distilled water 1:10), 1.4 ml 7.5% Na_2CO_3 . The samples were incubated in darkness, at room temperature for 30 min. The absorbance was measured at $\lambda = 765 \text{ nm}$ by spectrophotometer Shimadzu UV 1800. Standard curve based on known concentrations of gallic acid (GA) was used to calculate the amount of phenolic compounds as GA equivalents per dry weight ($\text{mg GA.g}^{-1} \text{ DW}$).

Total antioxidant activity analysis

The total antioxidant activity of each extract was measured according to PRIETO ET AL. (1999). Each sample contained 0.25 ml extract and 2.5 ml reagent solution (0.6 M H_2SO_4 , 28 mM CH_3COOK and 4 mM $(\text{NH}_4)_6\text{Mo}_7\text{O}_{24}$). The samples were incubated in a water bath for 90 min at 95°C . The reaction was stopped by placing the samples on ice. The absorbance was measured at $\lambda = 695 \text{ nm}$ by spectrophotometer Shimadzu UV 1800. The total antioxidant activity is calculated according to PRIETO ET AL. (1999) by multiplication with a coefficient from a standard curve with known concentrations of α -tocopherol and expressed as mM α -tocopherol per DW ($\text{mM.g}^{-1} \text{ DW}$).

Statistical analysis

The shown values are mean values of six to nine measurements (three extractions with two-three technical repetitions) and the related standard deviation. The *t*-test was applied for statistical evaluation with a threshold $P < 0.05$. For the correlation analysis, first, linear regression analysis was applied after checking the assumptions for normality and equality of the variances. Next, Pearson Product Moment Correlation coefficient (*r*) was calculated with $P = 0.05$ accepted as a level of significance. Data analysis was made by SigmaPlot software.

RESULTS

The maximal phenolics content of yarrow and thyme ranged in close limits (28.0–47.0 mgGA.gDW^{-1}). However, there were statistically significant differences between the material from the Rodopi Mts and Sofia regions. In yarrow, the phenolic content was higher in the plants from the Rodopi Mts in comparison to the plants from Sofia region (35.8 and 28.0 mgGA.gDW^{-1} , respectively). By contrast, in

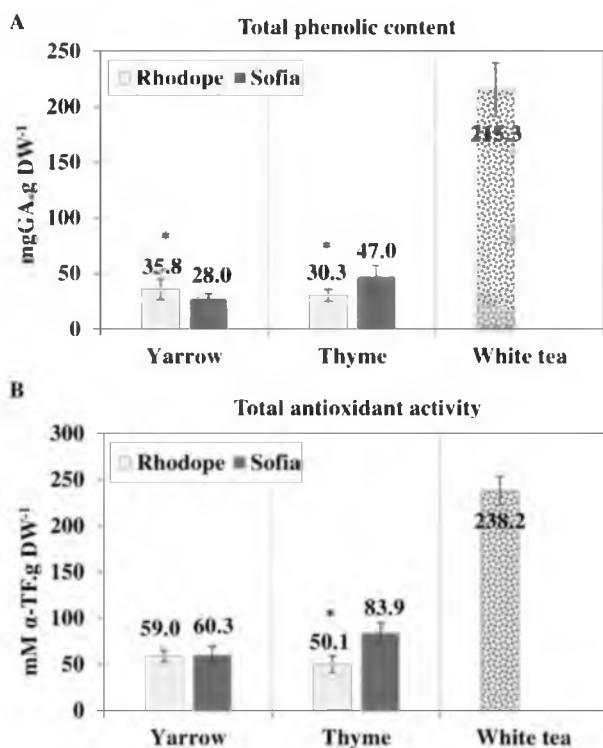


Fig. 1. Antioxidant potential of medicinal plants. Total content of phenolic compounds (A) and total antioxidant activity (B). Each variant from Rodopi Mts is compared to the one from Sofia region, and the presence of statistical difference is indicated with asterisks (* $P < 0.05$; $n > 6$).

four times higher (Fig. 1, 2).

Statistically significant Pearson correlation was established between the amount of phenolics and the antioxidant activity in thyme plants, from both studied regions ($r = 0.908$; $P < 0.001$) and in the white tea ($r = 0.999$; $P < 0.03$), as well. By contrast, in the case of yarrow such correlation was not established.

RESULTS

The maximal phenolics content of yarrow and thyme ranged in close limits (28.0–47.0 mgGA.gDW⁻¹). However, there were statistically significant differences between the material from the Rodopi Mts and Sofia regions. In yarrow, the phenolic content was higher in the plants from the Rodopi Mts in comparison to the plants from Sofia region (35.8 and 28.0 mgGA.gDW⁻¹, respectively). By contrast, in thyme, the phenolic content was higher in the plants obtained from Sofia region (30.3 and

47.0 mgGA.gDW⁻¹, respectively; Fig. 1A).

The maximal antioxidant activity in thyme was slightly higher than this in yarrow (83.9 and 60.3 mM α-TF.gDW⁻¹, respectively; Fig. 1 B). There was no difference between the antioxidant activity of the yarrow samples from the Rodopi Mts and Sofia, while in thyme a higher antioxidant potential was found in the material from Sofia (Fig. 1A, B).

The total phenolics content in the Chinese white tea was five to eight times higher in comparison to the studied yarrow and thyme material. Similarly, its antioxidant activity was

47.0 mgGA.gDW⁻¹, respectively; **Fig. 1A**).

The maximal antioxidant activity in thyme was slightly higher than this in yarrow (83.9 and 60.3 mM α -TF.gDW⁻¹, respectively; **Fig. 1B**). There was no

difference between the antioxidant activity of the yarrow samples from the Rodopi Mts and Sofia, while in thyme a higher antioxidant potential was found in the material from Sofia (**Fig. 1 A, B**).

The total phenolics content in the Chinese white tea was five to eight times higher in comparison to the studied yarrow and thyme material. Similarly, its antioxidant activity was four times higher (**Fig. 1 A, B**).

Statistically significant Pearson correlation was established between the amount of phenolics and the antioxidant activity in thyme plants, from both studied regions ($r=0.908$; $P < 0.001$) and in the white tea ($r=0.999$; $P < 0.03$), as well. By contrast, in the case of yarrow such correlation was not established.

DISCUSSION

There is a belief in Bulgaria that medicinal plants collected at sunrise after Saint John's Eve celebration (24th June) have great potential to cure and improve health. Our study did not reveal striking differences in the phenolic content and antioxidant activity of yarrow and thyme herbs collected in the natural habitats in the Rodopi Mts on 24th June and the same herbs obtained from random Bulgarian producers. Although the yarrow plants from both studied regions showed almost identical total antioxidant activities, their phenolic content differed. Since antioxidant capacity is not coming solely from the phenolics but could be due to the presence of some other phytochemicals (*e.g.* ascorbic acid, tocopherol, pigments, essential oils) or to the synergistic effects among them (SENGUL ET AL. 2009; KRAUJALIS ET AL. 2011), it could be suggested that some other compounds in the plants from Sofia region are capable of antioxidant activity, thus compensating the lower content of phenolics. This means that in yarrow, besides polyphenolics, there are additional metabolites to be explored that might have special beneficial effects on human health.

Our results are in accordance with some previous studies, which demonstrated the effect of the habitat on the antioxidant potential of yarrow. For example, considerable variation in accumulation of phenolic compounds among the flowers of *A. millefolium* L. from different localities was observed (BENETIS ET AL. 2008).

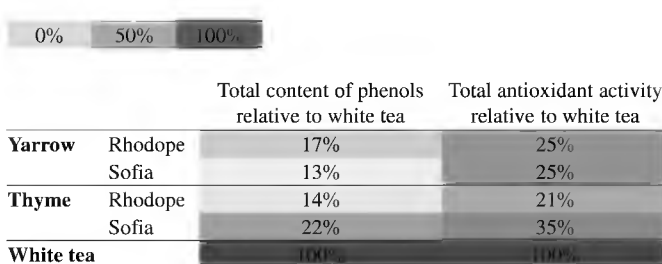


Fig. 2. Heat map data representation of the antioxidant potential of Bulgarian yarrow and thyme relative to the white tea.

In this study, the total amount of the identified phenolics in yarrow flowers from different populations varied from 13.290 to 27.947 mg.g⁻¹. Since the examined populations of *A. millefolium* were located in different regions of Lithuania within habitats with different environmental conditions (e.g. in microclimate, soil, ultraviolet radiation), it was concluded that the observed diversity could have a genetic basis, but it may be attributed also to the environmental differences. In Bulgaria, ROGOVA ET AL. (2015) performed similar screening of the antioxidant potential of the endemic species *A. thracica* Vel., grown *in vivo*, *in vitro* and *ex vitro* conditions, and reported lack of correlation between the phenolics content and the total antioxidant activity. Most of the studies on different *Achillea* species (including the endemic *A. thracica*) in Bulgaria were focused on their essential oil composition with revealing a dependence of the sesquiterpene lactone profile from the habitat or cultivation conditions (TODOROVA ET AL. 2000, 2004, 2007; YORDANOVA ET AL. 2017).

The comparison of the phenolics and antioxidant activity of all studied yarrow and thyme plants showed that they were several folds lower than in the Chinese white tea. As in other screens for the antioxidant potential of herbs, our work is an *in vitro* approach and the determined values could differ from those *in vivo* since polyphenols undergo extensive modification during digestion via conjugation in the intestinal cells and liver by sulphation, methylation, and glucuronidation (SETCHELL ET AL. 2003). Therefore, for revealing the real antioxidant potential of the investigated herbs, it would be more useful to use more antioxidant methods and to investigate the biological activities of the extracts from medicinal plants which can provide more detailed information about the specific roles of the metabolites (BADARINATH ET AL. 2010; MEKINIC ET AL. 2014). However, our results show the potential of the applied screening of total phenolics content and antioxidant activity as a fast approach to overview the general trend in the antioxidant potential and factors (such as genotype, habitat and storing conditions) that influence it in medicinal plant species used for infusions in the households.

ACKNOWLEDGEMENTS

The authors are grateful to the colleagues from specialty Molecular Biology in the Biological Faculty of Sofia University who provided technical support during the summer practice in Plant Physiology. We thank Assoc. Prof. ZHENYA YORDANOVA for the useful discussions regarding the work. The authors highly appreciate the critical comments and suggestions of the editors for improving the manuscript.

CONFLICT OF INTERESTS

The authors declare that there is no conflict of interests regarding the publication of this article.

AUTHOR CONTRIBUTIONS

N.G.G. performed experimental work; G.T.C. and M.K.Z. designed the experiments and interpreted the results; M.K.Z. wrote the manuscript.

References

- BADARINATH A. V., MALLIKARJUNA R. K., MADHU SUDHANA CHETTY C., RAMKANTH S., RAJAN T. V. S. & GNANAPRAKASH K. 2010. A review on *in-vitro* antioxidant methods: comparisons, correlations and considerations. - International Journal of PharmTech Research (2): 1276–1285.
- BENETIS R., RADUŠIENĖ J. & JANULIS V. 2008. Variability of phenolic compounds in flowers of *Achillea millefolium* wild populations in Lithuania. - Medicina (Kaunas) 44 (10): 775–781.
- BOUDET A. 2007. Evolution and current status of research in phenolic compounds. - Phytochemistry 68 (22-24): 2722–2735.
- BOUTAOUI N., ZAITER L., BENAYACHE F., BENAYACHE S., CARRADORI S., CESA S., GIUSTI A. M., CAMPESTRE C., MENGhini L., INNOSA D. & LOCATELLI M. 2018. Qualitative and quantitative phytochemical analysis of different extracts from *Thymus algeriensis* Aerial Parts. - Molecules 23 (2): pii: E463.
- DZIALO M., MIERZIAK J., KORZUN U., PREISNER M., SZOPA J. & KULMA A. 2016. The potential of plant phenolics in prevention and therapy of skin disorders. - International Journal of Molecular Science 17 (2): 160.
- EVSTATIEVA L., HARDALOVA R. & STOYANOVA K. 2007. Medicinal plants in Bulgaria: diversity, legislation, conservation and trade. – Phytologia Balcanica 13 (3): 415–427.
- FIEHN O. 2002. Metabolomics-the link between genotypes and phenotypes. - Plant Molecular Biology 48 (1-2): 155–171.
- GUPTA V. & SHARMA S. 2006. Plants as natural antioxidants. - Natural Product Radiance 5 (4): 326–334.
- JARIĆ S., MAČUKANOVIC-JOCIC M., DJURDJEVIĆ L., MITROVIĆ M., KOSTIĆ O., KARADŽIĆ B. & PAVLOVIĆ P. 2015. An ethnobotanical survey of traditionally used plants on Suva planina mountain (south-eastern Serbia). - Journal of Ethnopharmacology 175: 93–108.
- KAPCHINA-TOTEVA V., DIMITROVA M. A., STEFANOVA M., KOLEVA D., KOSTOV K., YORDANOVA ZH. P., STEFANOV D. & ZHIPONOVA M. K. 2014. Adaptive changes in photosynthetic performance and secondary metabolites during white dead nettle micropropagation. - Journal of Plant Physiology 171 (15): 1344–1353.
- KRATCHANOVA M., DENEV P., CIZ M., LOJEK A. & MIHAILOV A. 2010. Evaluation of antioxidant activity of medicinal plants containing polyphenol compounds. Comparison of two extraction systems. - Acta Biochimica Polonica 57 (2): 229–34.

- KRAUJALIS P., VENSKUTONIS P. R. & RAGAZINSKIENE O. 2011. Antioxidant activities and phenolic composition of extracts from *Nepeta* plant species. – In: Proceedings 6th Baltic Conference on Food Science and Technology Innovations for Food Science and Production - FOODBALT 2011, Jelgava, Latvia, 79-83.
- MEKINIC I. G., SKROZA D., LJUBENKOV I., KRSTULOVIC L., MOZINA S. S. & KATALINIC V. 2014. Phenolic acids profile, antioxidant and antibacterial activity of chamomile, common yarrow and immortelle (Asteraceae). - Natural Product Communications 9 (12): 1745-1748.
- MITHÖFER A. & BOLAND W. 2012. Plant defence against herbivores: chemical aspects. - Annual Review of Plant Biology 63: 431-450.
- PANDEY K. B. & RIZVI S. I. 2009. Plant polyphenols as dietary antioxidants in human health and disease. - Oxidative Medicine and Cellular Longevity 2 (5): 270-278.
- PEREIRA O. R., MACIAS R. I. R., PEREZ M. J., MARINC J. J. G. & CARDOSO S. M. 2013. Protective effects of phenolic constituents from *Cytisus multiflorus*, *Lamium album* L. and *Thymus citriodorus* on liver cells. - Journal of Functional Foods 5 (3): 1170-1179.
- PRIETO P., PINEDA M. & AGUILAR M. 1999. Spectrophotometric quantitation of antioxidant capacity through the formation of a phosphomolybdenum complex: Specific application to the determination of vitamin E1. - Analytical Biochemistry 269 (2): 337-341.
- ROGOVA M. A., DRAGOLOVA D. T., DIMITROVA M. A., YORDANOVA ZH. P., MANTOVSKA D., ZHIPONOVA M. & KAPCHINA-TOTEVA V. M. 2015. Micropropagation and ex situ conservation of *Achillea thracica* Velen. - Science & Technologies 5 (1), Medicine: 38-41.
- SENGUL M., YILDIZ H., GUNGOR N., CETIN B., ESER Z. & ERCISLI S. 2009. Total phenolic content, antioxidant and antimicrobial activities of some medicinal plants. - Pakistan Journal of Pharmaceutical Sciences 22 (1): 102-106.
- SETCHELL K. D., FAUGHNAN M. S., AVADES T., ZIMMER NECHEMIAS L., BROWN N. M., WOLFE B. E., BRASHEAR W. T., DESAI P., OLDFIELD M. F. & BOTTING N. P. 2003. Comparing the pharmacokinetics of daidzein and genistein with the use of ¹³C-labeled tracers in premenopausal women. - The American Journal of Clinical Nutrition 77 (2): 411-419.
- SHANNON E., JAISWAL A. K. & ABU-GHANNAM N. 2018. Polyphenolic content and antioxidant capacity of white, green, black, and herbal teas: a kinetic study. - Food Research 2 (1): 1-11.
- SINGLETON V. L., ORTHOFER R. & LAMUELA-RAVENTOS R.M. 1999. Analysis of total phenols and other oxidation substrates and antioxidants by means of Folin-Ciocalteu reagent. - Methods in Enzymology 299: 152-178.
- THRING S. A. T., HILI P. & NAUGHTON D. P. 2009. Anti-collagenase, anti-elastase and anti-oxidant activities of extracts from 21 plants. - BMC Complementary

and Alternative Medicine 9: 27.

- TODOROVA M., VOGLER B. & TSANKOVA E. 2000. Terpenoids from *Achillea setacea*. - Zeitschrift für Naturforschung C 55 (9-10): 840-842.
- TODOROVA M. N., TSANKOVA E. T. & MUSTAKEROVA E. I. 2004. Spirodepressolide: an unusual bis-norsesquiterpene lactone from *Achillea depressa* Janka. - Natural Product Research 18 (5): 461-464.
- TODOROVA M., TREDAFILOVA A., MIKHOVA B., VITKOVA A. & DUDDECK H. 2007. Chemotypes in *Achillea collina* based on sesquiterpene lactone profile. - Phytochemistry 68 (13): 1722-1730.
- YORDANOVA Z. P., ROGOVA M. A., ZHIPONOVA M. K., GEORGIEV M. I. & KAPCHINA-TOTEVA V. M. 2017. Comparative determination of the essential oil composition in Bulgarian endemic plant *Achillea thracica* Velen. during the process of *ex situ* conservation. - Phytochemistry Letters 20: 456-461.

Received 10 August 2018

Accepted 26 November 2018